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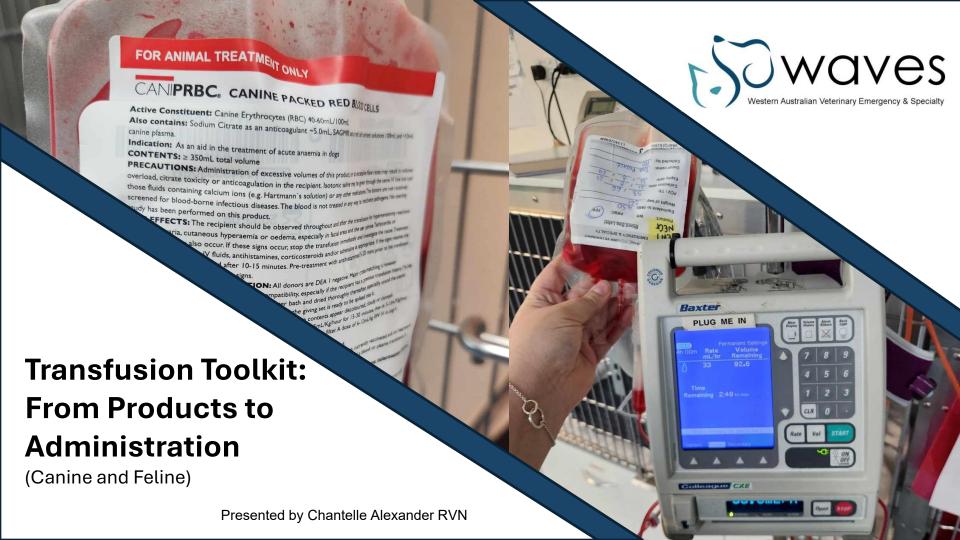












Let's Take the Confusion out of Transfusion...

By the end of today, you will be able to:

- Understand when and why to give a transfusion
- Discuss key blood products and when to use them
- > Escalate patient or lab changes to the vet
- Use a clear pre-transfusion checklist
- > Confidently administer and monitor a transfusion
- (Hopefully!) Recruit a few new blood donors to WAVES



Supportive VS Curative

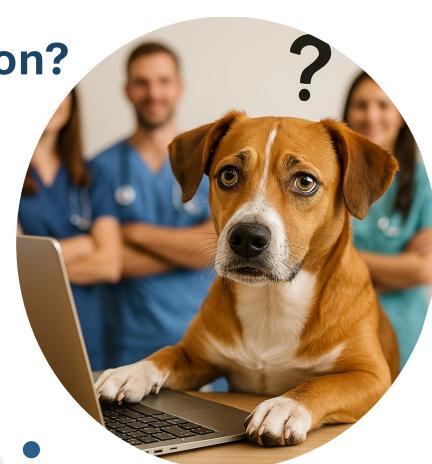


- > Transfusions are **supportive**, not curative
- Once consumed, the underlying disease can return
- ➤ Aim is to diagnose and treat the **primary cause**, not just the symptoms

Why Give a Transfusion?

Main goals:

- Restore blood volume
- Improve oxygen-carrying capacity
- Correct coagulopathies



**Increase Protein Levels?

Plasma is NOT an efficient fix for hypoalbuminaemia

- Albumin contributes ~80% to maintaining oncotic (colloid osmotic) pressure
- Helps with transport, buffering, and maintaining vascular volume
- Hypoalbuminaemia can cause GI stasis, ileus, intestinal oedema, nutrient malabsorption

Rule of thumb: 4.5 mL/kg plasma to raise albumin by 1 g/L.

So, a 30 kg dog with albumin of 10 g/L needs **1350 mL** whereas a 5kg patient would require **225ml** to reach a target of **20g/L**

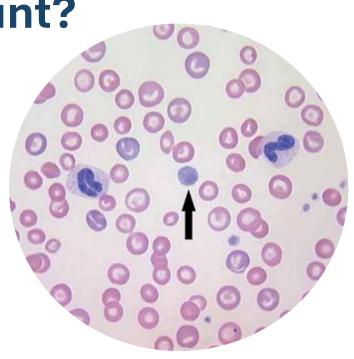
> Expensive and requires high volumes; increased risk of fluid overload



**Increase Platelet Count?

- ➤ Transfusions ≠ platelet support
- Only FWB replaces platelets
- Don't rely on transfusions; treat symptoms, not just the platelet count

1 mL/kg of fresh whole blood increases platelets by ~1 x 10°/L

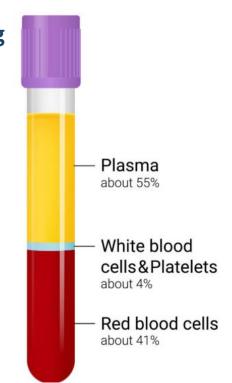


Fresh Whole Blood (FWB)

Fresh Whole Blood = RBCs + plasma + platelets + clotting factors

Use for patients who require:

- Anaemia with hypovolaemia
- Only way to transfuse functional platelets (can't be refrigerated)
- Rodenticide toxicity (RBCs + clotting factors)
 - 2 mL/kg typically increases PCV by ~1%



Stored Whole Blood (SWB)

Stored Whole Blood = RBCs + plasma + clotting factors

- Refrigeration stops platelets working
- Contains proteins and some clotting factors (many degrade over time, namely V, VIII)



Packed Red Blood Cells (PRBCs)

Why choose PRBC's over Whole Blood?

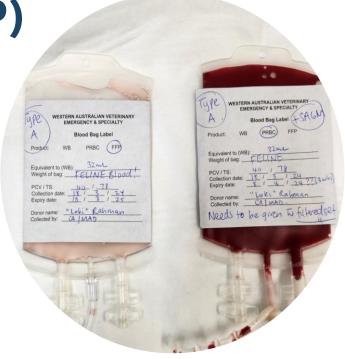
- Targeted therapy (give only what the patient needs)
- Less risk of volume overload
- > Fewer plasma-related reactions
- One donation = multiple patients
 - 1 mL/kg typically increases PCV by ~1%



Fresh Frozen Plasma (FFP) & Frozen Plasma (FP)

- Frozen Plasma: frozen >8 hrs after collection or FFP that's >1 year old
- ** Note: Naming of plasma products (FFP vs FP) may vary between countries and blood bank organisations.
- Replaces plasma proteins (clotting factors, albumin, immunoglobulins)

FFP dose: 10-20 mL/kg to treat coagulopathy



CaniPlas (Hyperimmune Plasma)

- From dogs with boosted or natural immunity (post-infection or repeat vaccination)
- May contain antibodies that help fight parvo**
- **Use is anecdotal no strong evidence it improves outcomes
- May help if albumin is low, but requires large volume (risk of fluid overland)





Cryoprecipitate (Honourable Mention)

- ➤ Plasma-derived, rich in fibrinogen, vWF, Factor VIII, XIII, fibronectin
- ➤ Made by slowly thawing FFP, then separating the protein-rich layer
- ➤ Small volume, targeted therapy for:
 - > vWD
 - > Haemophilia A
 - Hypofibrinogenaemia (DIC, severe bleeding)



Product	Targeted Applications
PRBCs	Anaemia (trauma, IMHA, chronic disease, surgical loss)
FFP	Coagulopathies (rodenticide toxicity, DIC, liver failure)
Frozen Plasma (FP)	Backup source for vitamin K-dependent factors (II, VII, IX, X)
Fresh Whole Blood (FWB)	Acute blood loss with need for red cells, clotting factors, and volume support
Cryoprecipitate	vWD, Haemophilia A, fibrinogen deficiency
Hyperimmune Plasma (Caniplas)	Passive immunity (parvovirus, sepsis)

Patient Parameters

- > Trends matter more than isolated values: one-off results can mislead; look at the bigger picture over time.
- > Tachycardia: Compensation to improve perfusion
- Tachypnoea: Compensation to increase oxygen delivery
- Pale mucous membranes: Reduced red cell mass/haemoglobin
- Weakness/lethargy: Inadequate oxygen to muscles/brain
- Cold extremities/weak pulses: Peripheral vasoconstriction to prioritise vital organs



Normal Values

Species	Heart Rate	Temperature	Respiratory Rate
Canine	60-140 bpm	37.5°C – 39.2°C	>40 bpm (at rest)
Feline	140-200 bpm	38.0°C – 39.2°C	>44 bpm (at rest)

Lab Values: Automated

Haematocrit (HCT) - Automated

A Haematology	6/5/25 11:29 am
■ W RBC	2.45 5.65 - 8.87 x10^12/L
M W Haematocrit	0.155 0.373 - 0.617 L/L
■ M Haemoglobin	47 131 - 205 g/L

- Calculated, not directly measured like manual PCV
- Uses RBC count × MCV, calculated using electrical signals and light sensors.
- > Influenced by:
 - > **Sample issues:** hemolysis, clots, lipemia, icterus, storage.
 - > **Analyzer issues:** calibration drift, misclassification, artifacts.
 - Patient factors: reticulocytosis, micro/macrocytosis, platelet clumping.

Lab Values- Manual

PCV (Packed Cell Volume) - Manual

- Manual equivalent of HCT
- Measured by spinning blood in a microhaematocrit tube
- Acute vs chronic anaemia (look at total solids & clinical signs)

Species	Packed Cell Volume (PCV)	Total Solids (TS)
Canine	35% - 55%	55-75 g/L
Feline	30% - 45%	60-80 g/L



Pre-Transfusion Checklist

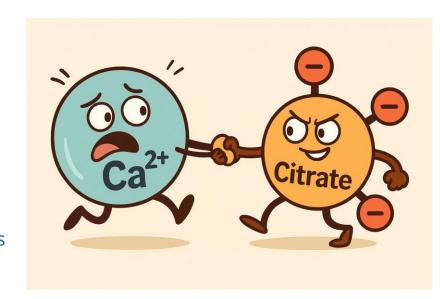
- Fluids & Medications Check
- Venous Access & Function
- **92** IVC Placement
- Blood Typing & Compatibility
- Crossmatching & Safety
- Patient Assessment & Baseline Vitals



Fluid Type	Compatible with BT?	Notes
0.9% NaCl (Saline)	✓ Yes	Gold standard
Plasma-Lyte 148	✓ Yes	Generally safe
CSL/LRS	▲ No (if same line)	Contains calcium – risk of precipitation
5% Dextrose	× No	Can cause red cell clumping (pseudoagglutination), haemolysis, and interfere with detecting true reactions.
0.45% NaCl	× No	Hypotonic – causes haemolysis in red cells, may impair efficacy of plasma
Hypertonic saline	× No	Causes crenation in red cells, can denature proteins or affect plasma integrity

Fluids and Medications

- > Avoid calcium-containing fluids (Hartmans, LRS)
- ➤ Ideally use **isotonic, calcium-free fluids**, such as:
 - 0.9% NaCl (Normal Saline)
 - Plasmalyte 148
- Change the fluid bag before transfusion
- **Prime all lines** (giving sets, T-pieces, extensions)
- **Avoid** IV antibiotics (metronidazole) and lipid emulsions



Venous Access

- ➤ Unwrap to check insertion site no kinks, swelling, or leakage
- ➤ Flush to confirm patency
- ➤ Check for signs of phlebitis:
 - Redness
 - Swelling
 - Heat
 - Pain/discomfort







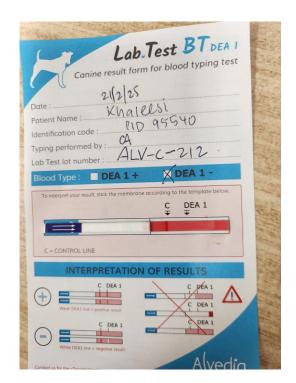


Placing an IVC

- **Clip wide** to minimise contamination
- **≻**Clean off debris and allow to dry
- **Wipe away excess chlorhex (esp. cats)
- >Avoid tape over site
- **≻Collect bloods** at placement
- **➤ Use largest gauge** suitable

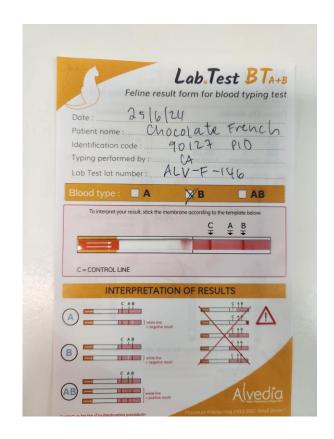
Blood Typing- Why it Matters

- > Type all transfusion patients prior to administration
- ➤ Help prevents life-threatening reactions
- Cats: Must be typed before a transfusion if blood type is unknown
- Dogs: Type once, especially for donors or patients expected to have repeat transfusions



Feline Blood Typing

- > Type A Most common
 - Weak anti-B antibodies
 - Can tolerate Type B in true emergencies (not ideal)
- > **Type B** Less common (30% of cats in Perth)
 - Strong anti-A antibodies → severe/fatal reactions with Type A
 - Common in Persians, Ragdolls, British/Devon/Cornish Rex
- > Type AB Rare
 - Type A recommended donor blood



Canine Blood Typing

- Many types, but focus on DEA 1
 - > **DEA 1 Positive** has the antigen
 - > **DEA 1 Negative** doesn't
- DEA 1 Negative: should only receive DEA 1 Negative
- > **DEA 1 Positive**: can receive either, but best to crossmatch
- Reactions usually occur with repeat transfusions



Species	Crossmatch Before First Transfusion?	Crossmatch Needed After Previous Transfusion?
Cats	Yes — ideally always	Yes — if >48 hrs since first transfusion
Dogs	• •	Yes — if >4 days since first transfusion

When to Crossmatch?

- Recommended even before any transfusion (canine & feline)
- Cats have natural antibodies → high reaction risk
- Some cats have anti-Mik antibodies → can cause transfusion reactions even when donor and recipient are the same blood type
- ➤ Ideally, always type + crossmatch whenever possible
- Can skip crossmatch if:
 - > Type is known
- Time-critical case (patient crashing)



Incidence of incompatible crossmatch results in dogs admitted to a veterinary teaching hospital with no history of prior red blood cell transfusion



Adesola Odunayo DVM, Kayode Garraway DVM, Barton W. Rohrba...

View More +

- 149 dogs with no prior transfusions
- All donors were DEA 1.1, 1.2, and 7 negative; DEA 4 positive (ideal donor dogs with low-risk blood types)
- > Still, **17% showed incompatibility** with 1 or 2 donors during crossmatching
- Dogs who were crossmatched had a **better rise in HCT** suggesting longer red cell survival.

Takeaway: Even "virgin" dogs can have unexpected antibodies → **crossmatch when possible**.

Major vs Minor Crossmatch

Major Crossmatch

- Tests recipient plasma vs donor RBCs
- Detects antibodies that could destroy donor red cells
- Most important and most commonly used in clinic

Minor Crossmatch

- Tests donor plasma vs recipient RBCs
- Useful when using fresh whole blood
- > Helps detect **donor antibodies** that might react with recipient cells

. Administering Blood Products Safely

- ➤ Check **product label** (type, expiry, volume)
- > Review fluids & CRIs
- ➤ Prepare equipment (monitors, infusion pumps, transfusion records, filtered giving sets)
- ➤ Assess the Patient: Look beyond numbers (mentation, behaviour, respiratory effort)
- ➤ Provide supportive care as needed (oxygen, antiemetics if nauseous, active warming etc)



Visual Inspection of Blood Product

- > No air bubbles
- > Colour: **rich red**, not dark/black
- > No cracks, leaks, or damaged seals
- > Must be **sterile**
- > If in doubt, don't use it tell the
- vet!



Blood Filtered Giving Sets

- Always use a blood filtered giving set for all blood products
- > Filters remove clots, debris & aggregates
- > Use an **infusion pump** where possible
 - Controls rate
 - > Tracks total volume
- Avoid infusion pumps unless designed for
- blood products





Infusion Rate & Monitoring

- Monitor patient every 5 mins until at target rate (HR, demeanor, temp, RR & RE)
- > Final rate depends on:
 - Product type
 - Vitals & disease status
 - Co-morbidities (cardiac disease)
- Aim to complete transfusion within 4 hours



Infusion Rate (cont'd)

- Start slow for first 15–30 mins to monitor for reaction
- Most patients aim for a rate of 10 mL/kg/hr
- Cardiac/overload risk: 4 mL/kg/hr
- Critical (shock/anaemia): As fast as needed
- Vet discretion applies! Keep monitoring & communicating



Blood Transfusion Record

Date: Patient name: Patient ID:

14/2/25 Lily Oldfield 97462 13/76

Blood product used: Donor name: Expiry:

PRBC Bonnie Britten 6/3/25

Pre-transfusion PCV/TS: Post-transfusion PCV/TS:

Volume administered: 100ml SMS Donor's Family Lick for yes

3 20 33 20/1/2 1/2 1/2 1/2	Time	Time Elapsed	Rate (ml/hr)	Demeanour	Temp	Pulse	Resp Rate	Comments
2 35 10 15 04R 38 L 110 52 01 2-40 15 20 02AK 38.4 12 144 2-45 20 30 02AK 38.4 112 44 2-55 30 30 02AK 38.4 110 40 3-25 1 hour 55 02AR 38.5 108 Pant 3-25 1h 30 55 02AR 38.6 84 32 1 mg.		0	5	RAK	38.3	160	Panl	
2-40 15 20 QAK 38.4 12 44 2-45 20 30 QAK 38.4 112 44 2-55 30 30 QAK 38.4 110 40 3-25 1 hour 55 QAR 38.5 108 Pant 3-35 1h 30 55 QAR 38.6 84 32 1104 L-25 2h 55 QAR 38.3 80 1104	2-30	5	10	C/A/C	38.4	130	Pant	
2-45 20 30 WAK 38.4 12 44 2-55 30 30 WAK 38.4 10 40 3-25 1 hour 55 WAR 78.5 108 Part 1 3-55 1h 30 55 WAR 38.6 84 32 1 mg.	235	10	15	0.44	38.4	115	52	On U3
2-55 30 30 00/10 384 116 40	7-40	15	20	RAK	38.k	1-7	44	
3-25 1 hour 55 QAR 38.5 108 Pant 3-55 1h 30 55 QAR 38.6 84 32 1 mg.	2.45	20	.30	WIK	38·4	112	44	
3-55 1h 30 55 QAR 38.0 84 32 1 mg.	2-55	30	30	CV/IC	35.4	110	40	
2h 55 QAIR 383 82 12h	3-25	1 hour	55	CAR	38.5	108	Punt	
	3-55	1h 30	55	QAK	38.0	34	32	1 my Hot
2h 30	4-25	2h	55	Q/1/8	38.3	3.	Pink	
		2h 30		14.				

Recognising Transfusion Reactions

Immunological (immune-mediated):

Febrile (most common):

 \triangleright Temp $\uparrow > 0.5$ °C, \uparrow HR, \uparrow RR, \uparrow RE

Allergic:

Hives, facial swelling, vomiting

Haemolytic (rare/severe):

Red urine, collapse, dull mentation

TRALI (lung injury):

Respiratory distress, hypoxia

Anaphylaxis:

Hypotension, collapse, vomiting



Recognising Transfusion Reactions

Non-immunological:

Fluid overload (TACO):

→ RR/effort, crackles, distended jugulars

Sepsis (contaminated product):

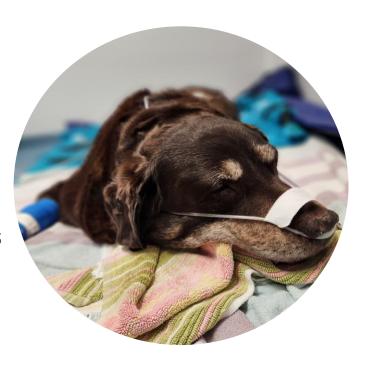
Sudden fever, hypotension, shock

Electrolyte disturbances (Ca, K, Mg):

Muscle tremors, arrhythmias

Embolism or massive transfusion complications

→ ↑ HR, ↑ RR, ↑ RE, hypotension, collapse, altered mentation, cyanosis or hypoxia, arrhythmias or ECG changes.



Suspected Transfusion Reaction – What Now?

- > Stop transfusion
- Alert the vet immediately, may be mild (WBCs/IgG) or serious (acute haemolysis)
- Continue to monitor, watch for improvement or decline
- May restart with:
 - Premedication
- Slower infusion rate



WAVES Blood Bank

- > Fear-free, positive visits to start with
- > Dogs, donation via cephalic vein
- Dogs, no sedation typically required
- Cats are screened for FIV, FeLV, Mycoplasma at first visit
- Cats are sedated for comfort, donation from a jugular vein
- Supported by IDEXX, Prime 100, Greenies





. Canine Blood Donors

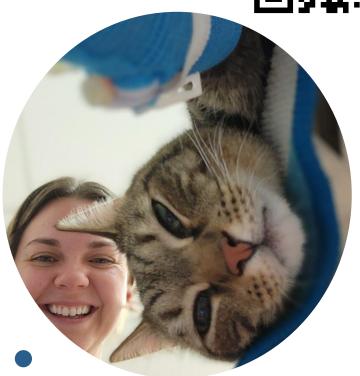
- > 1-7 years old
- > Over 28 kg lean body weight
- Up to date with vaccinations & parasite prevention
- Healthy with no signs of illness
- Never received a blood transfusion previously
- Never had a litter
- > Friendly, calm temperament



. Feline Blood Donors

- > 1-7 years old
- > Over 4 kg lean body weight
- > Indoor-only preferred
- Up to date with vaccinations & parasite prevention
- > Healthy with no signs of illness
- > Never received a blood transfusion
- Calm, handleable temperament





WAVES Plas Vacc PRBCs

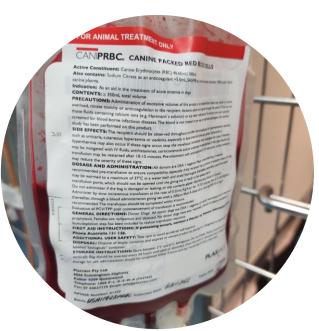


- CaniPRBC available only when blood bank stock allows
- > WAVES is a collection point only we don't hold stock
- First in, best dressed supply is limited
- No money exchanged between WAVES and your clinic
- Must be registered with Plasvacc as an authorised site
- PO must be completed (signed at collection or submitted electronically)
- Taxi collection possible PO must be received before dispatch
- We can only release the nearest-to-expiry unit
- No returns once it leaves WAVES, it's yours
- Need a filtered giving set? Let us know when requesting

Registered Clinics for PRBC Collection

Animal Emergency Care
Animalius Vet
Baldivis Vet Hospital
Bicton Vet Clinic
Claremont Vet Clinic
Comet Bay Vet Hospital
Duncraig Vet Hospital
Foothills Animal Hospital
Hamilton Hill Vet Hospital
Hammond Park Vet Clinic
Kalamunda Vet Hospital

Kwinana Vet Clinic
Malibu Vet Hospital
Millpoint Vet Centre
New Era Veterinary
Perth Vet Emergency
Ranford Vet Hospital
Sage Vets
Success Vets
Wanneroo Vet Hospital
Warnbro Vet Hospital



Questions??

Thank you for attending!!



References: August 1 2010, Andrea Oncken, DVM, DACVECC DVM360, 2008. Platelet disorders (Proceedings). [online] Available at: https://www.dvm360.com/view/platelet-disordersproceedings [Accessed 16 June 2025]. Giger, U., 2010. Transfusion Medicine—Do's and Don'ts. World Small Animal Veterinary Association World Congress Proceedings. Available at: https://www.vin.com/apputil/content/defaultadv1.aspx?id=4516258&pid=11310&print=1 [Accessed 16 June 2025]. Taylor, S., Spada, E., Callan, M.B., Korman, R., Leister, E., Steagall, P., Lobetti, R., Seth, M. and Tasker, S., 2021. 2021 ISFM consensus guidelines on the collection and administration of blood and blood products in cats. Journal of Feline Medicine and Surgery, 23(5), pp.410-432. https://doi.org/10.1177/1098612X211007071 Davidson, A. J., et al. (2013). Transfusion reactions in dogs and cats: 336 cases (2005–2010). Journal of the American Veterinary Medical Association, 243(5), 700–705. https://doi.org/10.2460/javma.243.5.700 Odunayo, A., Dodam, J.R., Bishop, M.A. and Campbell, V.L., 2017. Incidence of incompatible crossmatch results in dogs admitted to a veterinary teaching hospital with no history of prior red blood cell transfusion. Journal of the American Veterinary Medical Association, 250(3), pp.303-308. https://doi.org/10.2460/iavma.250.3.303 Mesa Sanchez, I. (ed.) (2025) Manual of Veterinary Transfusion Medicine: Principles, Techniques and Clinical Applications. Portugal: Animal Blood Bank. Norkus, C.P., 2018. Veterinary Transfusion Medicine. Hoboken, NJ: Wiley Blackwell.

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